
LIPID PEROXIDATION AND DNA DAMAGE AS BIOMARKERS OF POLLUTION INDUCED OXIDATIVE STRESS (OS) IN FISH

BHAWNA SRIVASTAVA, REDDY P.B

Abstract: The uses of maximum amount of agricultural and industrial chemicals are entering the aquatic environment and being taken up into tissues of aquatic organisms. Different types of pollutants induce a range of toxicity mechanisms, such as oxidative damage to membrane lipids, DNA, and proteins and changes to antioxidant enzymes. Free radical reactions and the production of toxic ROS are known to be responsible for a variety of oxidative damages leading to adverse health effects and diseases.

Though some literature is available on pollution induced oxidative stress but it is mostly based on mammalian studies. This assessment reviews current knowledge and advances in the understanding of such oxidative processes in aquatic organisms because of their sensitivity to oxidative pollutants and their potential for environmental toxicology studies. A search of literature was performed to collect the studies that measured the oxidative stress markers of pollution in fish. Studies were searched in Google scholar, Medline, Science direct, research gate, Pub Med, SCOPUS, Web of Science and other websites related to the subject from 1990 to May 2015.

Results indicate that escape of activated oxygen during active electron transport is the main source of ROS in man and higher animals. Another possibility is that multiple redox-active flavoproteins also contribute a small fraction to the overall production of oxidants under normal conditions. To maintain proper cellular homeostasis a balance must be struck between the production and consumption of ROS. It is concluded that measurements of lipid peroxidation and DNA damage both in nucleus and mitochondria can be used as potential contenders for general biomarkers of oxidative stress. However, these markers may be noticeable differently in the field than in results found in laboratory studies. Hence a multifaceted approach should be taken in field studies.

Keywords: Antioxidant, Environmental pollution, Fish, Oxidative Stress, Xenobiotics.

1. Introduction: Living organisms encounter an array of stresses during their constant interaction with environment [1]. Anthropogenic impacts on aquatic ecosystems are prevalent in developed and developing countries. Over extraction of freshwater, mainly for industries and agriculture has lead to major deprivation of rivers, lakes and aquifers. Environmentally stimulated stresses often activate the endogenous production of reactive oxygen species (ROS), most of which are generated as derivatives of mitochondrial respiration. Therefore regular exposure to toxicants may enhance ROS-mediated oxidative damage. More amount of agricultural and industrial wastes enter aquatic environment and being taken up by aquatic organisms induce multiple changes. Some of them directly enhance ROS formation whereas others act indirectly [2]. Oxygen is essential for efficient energy production in all aerobic organisms. Reactive oxidative species (ROS) are formed as natural derivatives of the normal metabolism of oxygen and have important roles in cell signaling and homeostasis [3], [4]. But, due to environmental stress (UV, heavy metal pollution, pathogens, heat exposure) ROS levels can increase significantly and induce oxidative stress (OS) which may affect major damage to cell and tissue organization and causes neurodegenerative disorders

in man and other animals [5], [6], [7], [8]. In human beings, oxidative stress is thought to be connected in the development various diseases [5], [7] including cancer [9] and myocardial infarction [10]. On the other hand, reactive oxygen species can be helpful, as they are used by the immune system as a means to attack and kill pathogens [11]. Temporary oxidative stress with short duration may also be vital in prevention of aging by induction of a process named mitohormesis [12].

Therefore, aquatic ecosystem is a sink for many ecological pollutants which can be taken up by aquatic organisms leading to disturbing of prooxidant/antioxidant balance in fish [13],[14], [15]. So only the direct determining of the oxidative stress response in biological systems has become the most adequate tool for early warning in ecotoxicology studies [13].

2. Methodology: A systematic search of Pub Med, Scopus databases, Med line, Google Scholar, Science direct, Researchgate and the reference lists of all included studies and major relevant review papers was performed to obtain the data. To find out the appropriate articles, Pub Med was searched using the key words prooxidants, antioxidants, aquatic organisms and ROS in various combinations. Case-control studies with fish subjects were considered for

inclusion. The articles selected were published in English between January 1985 and December 2015.

3. Results and Discussion: Living beings have distinctive systems for defending themselves against the detrimental effects of activated ROS. Oxidative stress (OS) can result in oxidative damage to lipids, proteins, carbohydrates, and nucleic acids and is considered to happen when there is a difference in the prooxidant-to-antioxidant ratio in biological system. In majority cases, the anomalous production of ROS can result in major damage to cell structure [16]. The indication of exact symptoms known to occur particularly at oxidative stress is lipid peroxidation (membrane damage) DNA damage and accumulation of lipofuscin pigments were found in many aquatic animals exposed to contaminants [17]. Therefore, examining the change in activity of antioxidant enzymes such as SOD, CAT, and GPx is believed as a valuable method of indicating the oxidative stress. In recent times, degree of difference expression of the genes encoding these enzymes has also been used to identify biological toxicity and/or to monitor the impact of chemical pollutants [18], [19].

Biomarkers of Lipid Peroxidation: Usually lipid peroxidation (LPO) or oxidation of poly unsaturated fatty acids is measured as a level of thiobarbituric acid reactive substances (TBARS). It has been employed most often to analyze the effect of pollutants [14], [15], [20], [21]. Lipid peroxidation (LPO) is the oxidative degradation of lipids and occurs due to oxidative imbalance. It is the process in which free radicals take electrons from the lipids in cell membranes and causes changes in the fluidity and permeability of cell membranes and impairs the activity of membrane-bound enzymes, resulting in cell damage [22]. Lipid peroxidation also leads to the production various aldehydes like malondialdehyde (MDA), 4-hydroxynonenal (HNE), and thiobarbituric acid reactive substances (TBARS). The elevated LPO in fish from heavily polluted field sites was observed [23]. For instance, Farombi, et al. [35] demonstrated high level of TBARS in the African catfish (*Clarias gariepinus*) from the Ogun River of Nigeria, located close to major industries. TBARS levels were significantly higher in the liver, kidney, gills and heart by 177%, 102%, 168% and 71% respectively compared to control fish. Similarly, higher levels of LPO products were found in the blood of three cichlid fish species (*Oreochromis niloticus*, *Tilapia rendalli*, and *Geophagus brasiliensis*) from the site of heavy metal pollution. [24] (Stoliar, O.B. and Lushchak, V.I., 2012). In another experiment Dorval et al, [25] showed elevated level of hepatic LPO products in white sucker (*Catostomus commersoni*) from the river sites that contaminated by agricultural chemicals in Québec (Canada). The killifish (*Fundulus heteroclitus*) inhabiting of the Elizabeth River also

exhibited higher LPO as compared to the control fish [26]. Variations in the level of TBARS in a liver of common carp (*Cyprinus carpio*) were also noticed between fish from rural and industrial sites in Western Ukraine[27]

Several studies also demonstrate that TBARS level in fish exposed to various pollutants is significantly higher than in controls. Nwani, C.D et al [28] found elevated level of lipid peroxidation in the liver of *C. punctatus* in response to the exposure to herbicide atrazine due to increased production of ROS. Increased ROS production may be associated with the metabolism of herbicide leading to the peroxidation of membrane lipids of the hepatocytes. Likewise, earlier investigations have reported the induction of lipid peroxidation by pesticides such as deltamethrin [29] alachlor,[30] malathion, [31], endosulfan [32], butachlor [23], pollutants [33], toxic metals [34], sewage [35] and effluent in fish [36].

But in contrast, But, in contrast elevated levels of lipid peroxidation was not observed in the brain and liver of goldfish *Crassius auratus* exposed to sub lethal concentration of Roundup® [37]. Other case study by Pandey et al [38] shown the absence of differences in TBARS concentration in gills, kidney and liver tissues of fish (*Wallago attu* (Bl. & Schn.) from polluted and non polluted areas. Similar results were found in the hepatopancreas of carp from polluted site [39]. Even though differences observed in the activities of antioxidant the intensity of LPO was the same in the fish from the both sites, signifying a stronger antioxidant capacity of this organ. Likewise, TBARS did not show inter-site differences in eelpout (*Z. viviparous*) [40]. In the same way, despite the variations in the antioxidant enzyme activities, there was no significant difference in TBARS concentrations in the liver of labrid fish (*Coris julis*) [41]. The diverse responses in antioxidant enzymes level and TBARS concentrations may be due differences in the species, the time and duration of exposure, type and concentration of stressors.

Lipofuscins pigments are the end-products of LPO can be gathered in lysosomes as insoluble granules. The sign of these pigments in the lysosomes also can be used for the evaluation of the level of membrane LPO [42]. In our opinion, the evaluation of lipofuscin levels as more suitable characteristic of damage to lipids than TBARS. However, the corresponding studies with fish in the field are limited and connected exclusively histological studies which may not permit to assess the oxidative stress response exactly. For example, the histopathological investigations in freshwater fish, i.e. largemouth bass (*Micropterus salmoides*) and redbreast sunfish (*Lepomis auratus*), showed the marks of lipofuscin pigment accumulation in polluted sites [43].

DNA damage: In aerobic cells, the uncontrolled reactive oxygen species (ROS) which are generated as a by-product of mitochondrial activity can cause severe damage to cellular macromolecules, particularly the DNA [44]. It is very well known that there are close correlation between oxidative stress (OS) and DNA damage [45]. Pollution induced ROS can provoke oxidative damage of DNA, including strand breaks and base and nucleotide modifications, particularly in sequences with high guanosine content [46]. However, the application of these methods in fish is inadequate. In a study by Rodriguez-Ariza et al [47] found that 8-oxodG determination in chromosomal DNA was a potentially useful biomarker of oxidative stress caused by urban and industrial effluents in a fish (*Sparus aurata*). Jin, Y et al [48] have studied the effects of cypermethrin exposure on the induction of hepatic oxidative stress and DNA damage in adult zebra fish and found that even low concentration of pesticide can cause heavy DNA damage and gene expression. Similarly, Patel et al [49] reported cypermethrin induced DNA damage in the brain, liver, and kidney, of mice. A significant dose-dependent increase in DNA damage was observed in

the brain cells and mid gut of *Drosophila melanogaster* exposed to low concentrations of cypermethrin [50].

4. Conclusions: The above outcome of this study implies that oxidative stress (OS) induced by various pollutants including heavy metals is a central issue in aquatic ecosystems. It also provides an insight into the role of reactive species in pollution induced toxicity. For that reason fish can be used as model organism for the study of pollution induced oxidative stress. However, the species specific forms of biomarkers and mechanisms of their action still need to be investigated. Chemically mediated pathways can affect antioxidant responses at different levels including transcriptional, translational and catalytic functions. In aquatic organisms such mechanisms remain largely unexplored in environmental monitoring programmes. Besides, molecular responses of antioxidants are often not analogous by expected biochemical changes or cellular effects. So, care is required when interpreting the outcomes of ecological pollutants. Finally it is concluded that cell stress biomarkers may serve as important tools for biomonitoring and development of risk assessment protocols.

References:

- Reddy P.B. and Baghel B.S. 2012. Impact of Industrial waste water on the Chambal River and Biomarker responses in fish due to pollution at Nagda. M.P. India. DAV Int. J. Sci. 1(1): 86-91.
- Stoliar, O.B. and Lushchak, V.I., 2012. Environmental pollution and oxidative stress in fish. INTECH Open Access Publisher.
- Devasagayam, T.P.A., Tilak, J.C., Boloor, K.K., Sane, K.S., Ghaskadbi, S.S. and Lele, R.D., 2004. Free radicals and antioxidants in human health: current status and future prospects. Japi, 52(794804), p.4.
- Naito, Aoi, W.Y., Mizushima, K., Takanami, Y., Kawai, Y., Ichikawa, H. and Yoshikawa, T., 2010. The micro RNA miR-696 regulates PGC-1 α in mouse skeletal muscle in response to physical activity. American Journal of Physiology-Endocrinology and Metabolism, 298(4), pp.E799-E806.
- Singh, N., Dhalla, A.K., Seneviratne, C., Singal, P.K. 1995. Oxidative stress and heart failure". Molecular and Cellular Biochemistry 147 (1): 77-81.
- Pohanka, M. 2013. Alzheimer's disease and oxidative stress: a review". Current Medicinal Chemistry, 21 (3): 356-364.
- Pohanka, M. 2013. Role of oxidative stress in infectious diseases. A review." Folia Microbiologica, 584 (6): 503-513.
- Kala Chandra, Ali Syed Salman, Abed Mohd, Rajpoot Sweetey, Khan Najam Ali. 2015. Protection against FCA Induced Oxidative Stress Induced DNA Damage as a Model of Arthritis and in vitro Anti-arthritic Potential of Costus speciosus Rhizome Extract. www.ijppr.com International Journal of Pharmacognosy and Phytochemical Research, 7(2); 383-389.
- Halliwell, B. 2007. Oxidative stress and cancer: have we moved forward?. Biochemical Journal, 401(1), pp.1-11. Halliwell, B., 2007. Oxidative stress and cancer: have we moved forward?. Biochemical Journal, 401(1), pp.1-11.
- Ramond A, Godin-Ribuot D, Ribuo C, Totoson P, Koritchneva I, Cachot S, Levy P, Joyeux-Faure M. 2011. Oxidative stress mediates cardiac infarction aggravation induced by intermittent hypoxia." Fundam Clin Pharmacol. 27 (3): 252-261.
- Torres, M.A., Jones, J.D. and Dangl, J.L., 2006. Reactive oxygen species signaling in response to pathogens. Plant physiology, 141(2), pp.373-378.
- Andersson, V., 2014. Links between Aging and Proteostasis Decline in *Saccharomyces cerevisiae*.
- Valavanidis, A., Vlahogianni, T., Dassenakis, M. and Scoullou, M., 2006. Molecular biomarkers of oxidative stress in aquatic organisms in relation to toxic environmental pollutants. Ecotoxicology and environmental safety, 64(2), pp.178-189.

14. Livingstone, D.R., 2003. Oxidative stress in aquatic organisms in relation to pollution and aquaculture. *Revue de Medecine Veterinaire*, 154(6), pp.427-430.
15. Lushchak, V.I., 2011. Environmentally induced oxidative stress in aquatic animals. *Aquatic Toxicology*, 101(1), pp.13-30.
16. Barzilai, A. and Yamamoto, K.I., 2004. DNA damage responses to oxidative stress. *DNA repair*, 3(8), pp.1109-1115.
17. Winston, G.W., 1991. Oxidants and antioxidants in aquatic animals. *Comp. Biochem. Physiol. C* 100, 173-176.
18. Sheader, D.L., Williams, T.D., Lyons, B.P., Chipman, J.K., 2006. Oxidative stress response of European flounder (*Platichthys flesus*) to cadmium determined by a custom cDNA microarray. *Mar. Environ. Res.* 62, 33-44.
19. Roh, J.Y., Lee, J. and Choi, J., 2006. Assessment of stress-related gene expression in the heavy metal-exposed nematode *Caenorhabditis elegans*: a potential biomarker for metal-induced toxicity monitoring and environmental risk assessment. *Environmental Toxicology and Chemistry*, 25(11), pp.2946-2956.
20. Lushchak, V.I. and Bagnyukova, T.V., 2006. Effects of different environmental oxygen levels on free radical processes in fish. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 144(3), pp.283-289.
21. Lushchak, V., Kubrak, O.I., Nykorak, M.Z., Storey, K.B. and Lushchak, V.I., 2008. The effect of potassium dichromate on free radical processes in goldfish: possible protective role of glutathione. *Aquatic Toxicology*, 87(2), pp.108-114.
22. Grim, J.M., Semones, M.C., Kuhn, D.E., Kriska, T., Keszler, A. and Crockett, E.L., 2015. Products of lipid peroxidation, but not membrane susceptibility to oxidative damage, are conserved in skeletal muscle following temperature acclimation. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 308(5), pp.R439-R448.
23. Farombi, E.O., Adelowo, O.A. and Ajimoko, Y.R., 2007. Biomarkers of oxidative stress and heavy metal levels as indicators of environmental pollution in African cat fish (*Clarias gariepinus*) from Nigeria Ogun River. *International Journal of Environmental Research and Public Health*, 4(2), pp.158-165.
24. Stoliar, O.B. and Lushchak, V.I., 2012. Environmental pollution and oxidative stress in fish. INTECH Open Access Publisher.
25. Dorval, J., Leblond, V., Deblois, C. and Hontela, A., 2005. Oxidative stress and endocrine endpoints in white sucker (*Catostomus commersoni*) from a river impacted by agricultural chemicals. *Environmental Toxicology and Chemistry*, 24(5), pp.1273-1280.
26. Bacanskas, L.R., Whitaker, J. and Di Giulio, R.T., 2004. Oxidative stress in two populations of killifish (*Fundulus heteroclitus*) with differing contaminant exposure histories. *Marine environmental research*, 58(2), pp.597-601.
27. Falfushynska, H.I., Gnatyshyna, L.L., Stoliar, O.B. and Nam, Y.K., 2011. Various responses to copper and manganese exposure of *Carassius auratus gibelio* from two populations. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 154(3), pp.242-253.
28. Nwani, C.D., Lakra, W.S., Nagpure, N.S., Kumar, R., Kushwaha, B. and Srivastava, S.K., 2010. Toxicity of the herbicide atrazine: effects on lipid peroxidation and activities of antioxidant enzymes in the freshwater fish *Channa punctatus* (Bloch). *International Journal of Environmental Research and Public Health*, 7(8), pp.3298-3312.
29. Atif, F., Parvez, S., Pandey, S., Ali, M., Kaur, M., Rehman, H., Khan, H.A. and Raisuddin, S., 2005. Modulatory effect of cadmium exposure on deltamethrin-induced oxidative stress in *Channa punctata* Bloch. *Archives of environmental contamination and toxicology*, 49(3), pp.371-377.
30. Peebua L.P., Kosiyachinda P., Pokethitiyook P., Kruatrachue M. 2007. Evaluation of alachlor herbicide impacts on Nile tilapia (*Oreochromis niloticus*) using biochemical biomarkers - *Bull. Environ. Contam. Toxicol.* 78: 138-141.
31. Monteiro, D.A., Rantin, F.T. and Kalinin, A.L., 2010. Inorganic mercury exposure: toxicological effects, oxidative stress biomarkers and bioaccumulation in the tropical freshwater fish *matrinxã*, *Brycon amazonicus* (Spix and Agassiz, 1829). *Ecotoxicology*, 19(1), pp.105-123.
32. Pandey, S., Ahmad, I., Parvez, S., Bin-Hafeez, B., Haque, R. and Raisuddin, S., 2001. Effect of endosulfan on antioxidants of freshwater fish *Channa punctatus* Bloch: 1. Protection against lipid peroxidation in liver by copper preexposure. *Archives of environmental contamination and toxicology*, 41(3), pp.345-352.
33. Fatima, M., Ahmad, I., Sayeed, I., Athar, M. and Raisuddin, S., 2000. Pollutant-induced over-activation of phagocytes is concomitantly associated with peroxidative damage in fish tissues. *Aquatic Toxicology*, 49(4), pp.243-250.
34. Ercal, N., Gurer-Orhan, H. and Aykin-Burns, N., 2001. Toxic metals and oxidative stress part I: mechanisms involved in metal-induced oxidative damage. *Current topics in medicinal chemistry*, 1(6), pp.529-539.
35. Guiloski, I.C., Ribas, J.L.C., da Silva Pereira, L., Neves, A.P.P. and de Assis, H.C.S., 2015. Effects of trophic exposure to dexamethasone and

- diclofenac in freshwater fish. *Ecotoxicology and environmental safety*, 114, pp.204-211.
36. Javed, M., Usmani, N., Ahmad, I. and Ahmad, M., 2015. Studies on the oxidative stress and gill histopathology in *Channa punctatus* of the canal receiving heavy metal-loaded effluent of Kasimpur Thermal Power Plant. *Environmental monitoring and assessment*, 187(1), pp.1-11.
37. Lushchak, V., Kubrak, O.I., Storey, J.M., Storey, K.B. and Lushchak, V.I., 2009. Low toxic herbicide Roundup induces mild oxidative stress in goldfish tissues. *Chemosphere*, 76(7), pp.932-937.
38. Pandey, S., Parvez, S., Sayeed, I., Haque, R., Bin-Hafeez, B. and Raisuddin, S., 2003. Biomarkers of oxidative stress: a comparative study of river Yamuna fish *Wallago attu* (Bl. & Schn.). *Science of the total environment*, 309(1), pp.105-115.
39. Huang, D.J., Zhang, Y.M., Song, G., Long, J., Liu, J.H. and Ji, W.H., 2007. Contaminants-induced oxidative damage on the carp *Cyprinus carpio* collected from the upper Yellow River, China. *Environmental monitoring and assessment*, 128, (1-3), pp.483-488.
40. Almroth B.C., Albertsson, E. and Förlin L.S.J. 2008. Oxidative stress, evident in antioxidant defenses and damage products, in rainbow trout caged outside a sewage treatment plant. *Ecotoxicol Environ Safety*.
41. Sureda, A., Box, A., Enseñat, M., Alou, E., Tauler, P., Deudero, S. and Pons, A., 2006. Enzymatic antioxidant response of a labrid fish (*Coris julis*) liver to environmental caulerpenyne. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 144(2), pp.191-196.
42. Viarengo, A., Canesi, L., Pertica, M., Poli, G., Moore, M.N. and Orunesu, M., 1990. Heavy metal effects on lipid peroxidation in the tissues of *Mytilus galloprovincialis* lam. *Comparative Biochemistry and Physiology Part C: Comparative Pharmacology*, 97,(1), pp.37-42.
43. Brunk, U.T. and Terman, A., 2002. Lipofuscin: mechanisms of age-related accumulation and influence on cell function. *Free Radical Biology and Medicine*, 33(5), pp.611-619.
44. Barzilai, A., Yamamoto, K.I., 2004. DNA damage responses to oxidative stress. *DNA Repair* 3, 1109-1115.
45. Kasai, H., 1997. Analysis of a form of oxidative DNA damage, 8-hydroxy-2'-deoxyguanosine, as a marker of cellular oxidative stress during carcinogenesis. *Mutation Research/Reviews in Mutation Research*, 387(3), pp.147-163.
46. Bennett, M.R., 2001. Reactive oxygen species and death oxidative DNA damage in atherosclerosis. *Circulation Research*, 88(7), pp.648-650.
47. Rodriguez-Ariza, A., Alhama, J., Diaz-Mendez, F.M. and Lopez-Barea, J., 1999. Content of 8-oxodG in chromosomal DNA of *Sparus aurata* fish as biomarker of oxidative stress and environmental pollution. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 438(2), pp.97-107.
48. Jin, Y., Zheng, S., Pu, Y., Shu, L., Sun, L., Liu, W. and Fu, Z., 2011. Cypermethrin has the potential to induce hepatic oxidative stress, DNA damage and apoptosis in adult zebra fish (*Danio rerio*). *Chemosphere*, 82(3), pp.398-404.
49. Patel, S., Pandey, A.K., Bajpayee, M., Parmar, D. and Dhawan, A., 2006. Cypermethrin-induced DNA damage in organs and tissues of the mouse: evidence from the comet assay. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 607(2), pp.176-183.
50. Mukhopadhyay, I., Chowdhuri, D.K., Bajpayee, M. and Dhawan, A., 2004. Evaluation of in vivo genotoxicity of cypermethrin in *Drosophila melanogaster* using the alkaline Comet assay. *Mutagenesis*, 19, (2), pp.85-90.

Bhawna Srivastava, Reddy, P.B

Department of Zoology, DAV College, Kanpur

PG Department of Zoology, Government PG College, Ratlam. M.P. India.